



Combined genotypic and phenotypic AST through gene expression profiling

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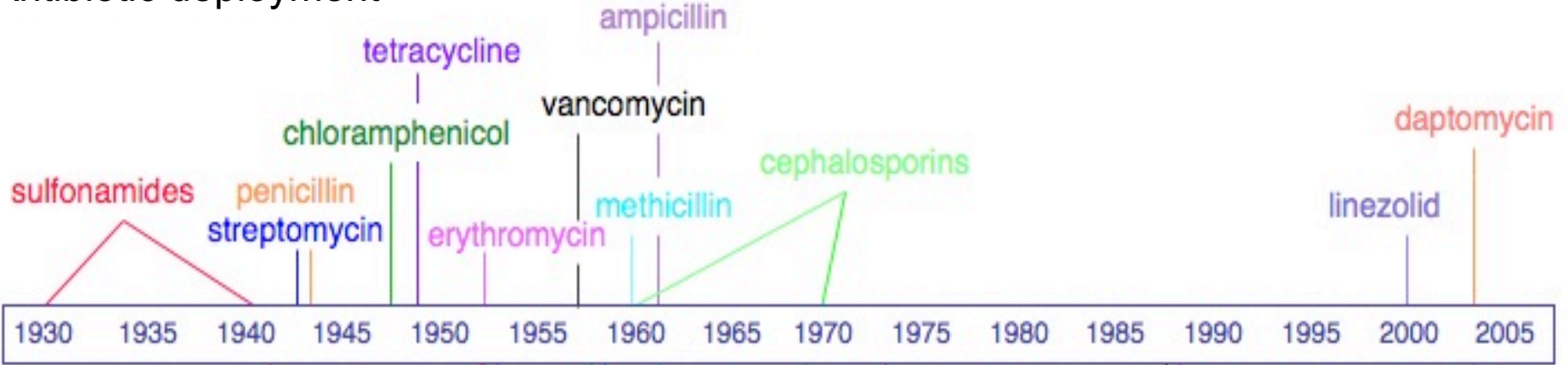
 @roby_bhatt

Disclosures

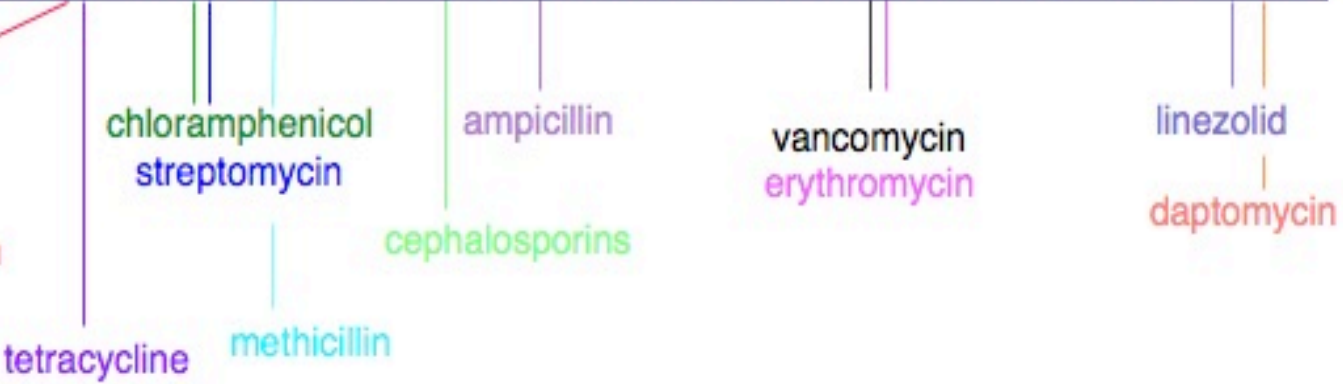
- None
- Some of this work was supported by an R01 from NIH/NIAID, awarded as an academia-industry partnership with NanoString

Antibiotics: a scarce societal good

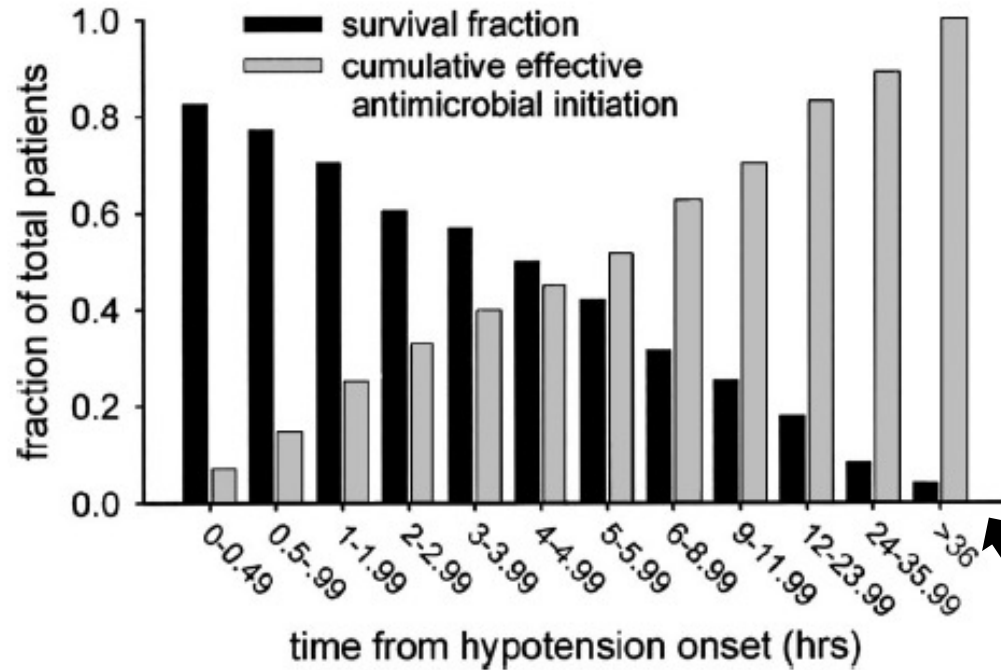
Antibiotic deployment



Emergence of antibiotic resistance

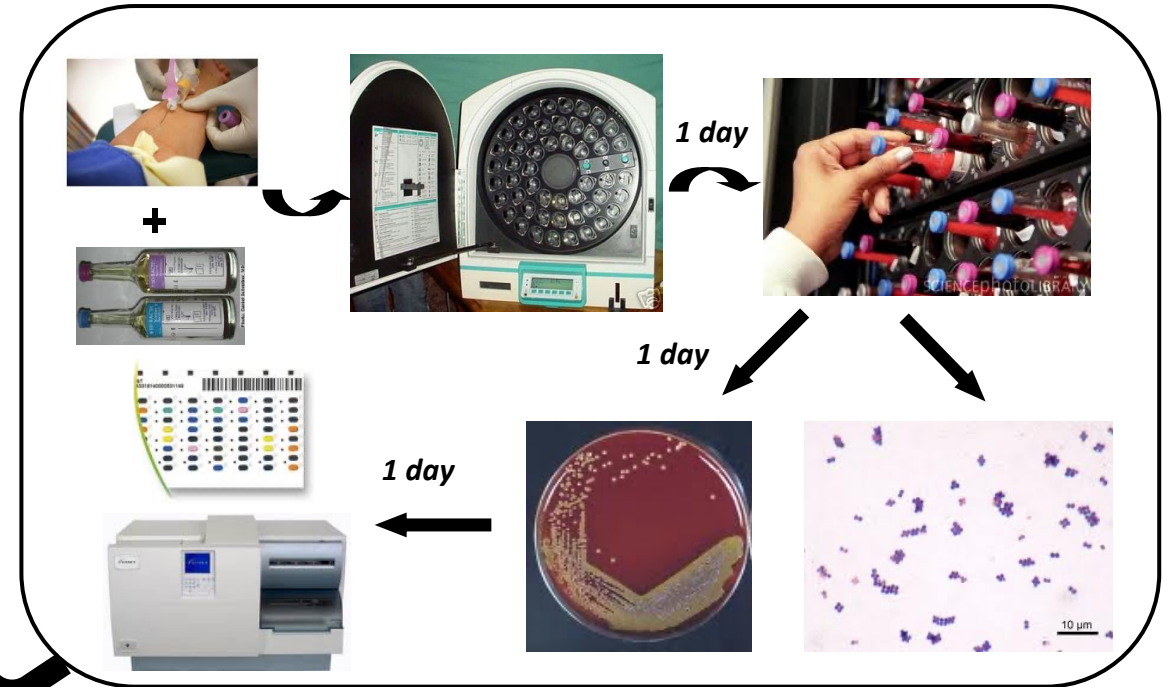


Current bacterial diagnostics are too slow to meet the clinical mandate



Kumar A et al, *Crit Care Med* 2006;34:1589

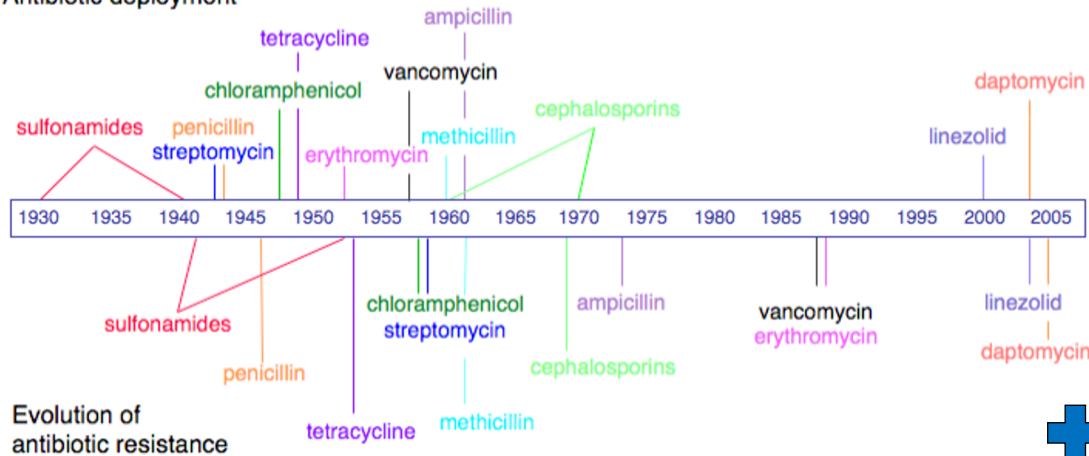
2-3 days



- Every hour's delay to giving appropriate antibiotics increases mortality from severe sepsis by 7%
- Best-case scenario for growth-based AST is 2-3 days from presentation

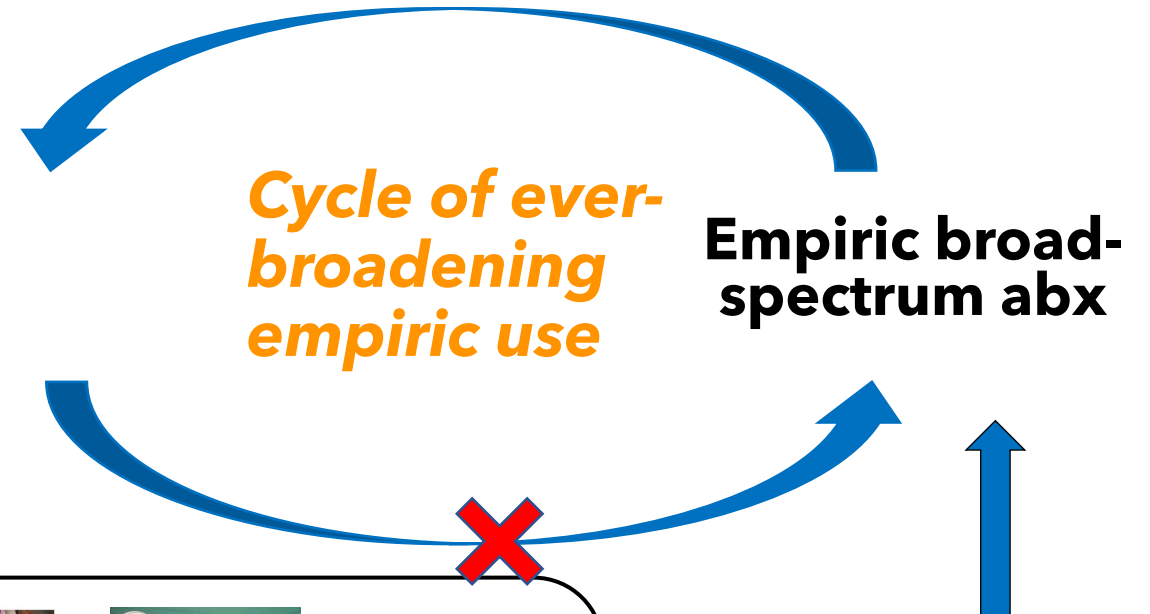
This combination leads to empiric broad-spectrum antibiotic use, feeding the cycle

Antibiotic deployment



Evolution of antibiotic resistance

1.0 — survival fraction



- To break the cycle of escalating antibiotic use, diagnostics must be **fast** AND **accurate**
 - Ensure efficient deployment of scarce antibiotics
 - Potential to **resolve tension** between individual and society
 - Consider Oncology: from poisons to targeted therapy with molecular diagnostics

Genomics and AST

- How can we find evidence of **resistance** in genomic data?
 - Hydrolases, acetylases, efflux pumps
 - May even work better than phenotype in some cases, eg carbapenemase producers¹
 - Target site mutations? Gene inactivations?
 - Changes in expression or copy number??

¹Humphries RM, *J Clin Micro* 2019

Genomics and AST

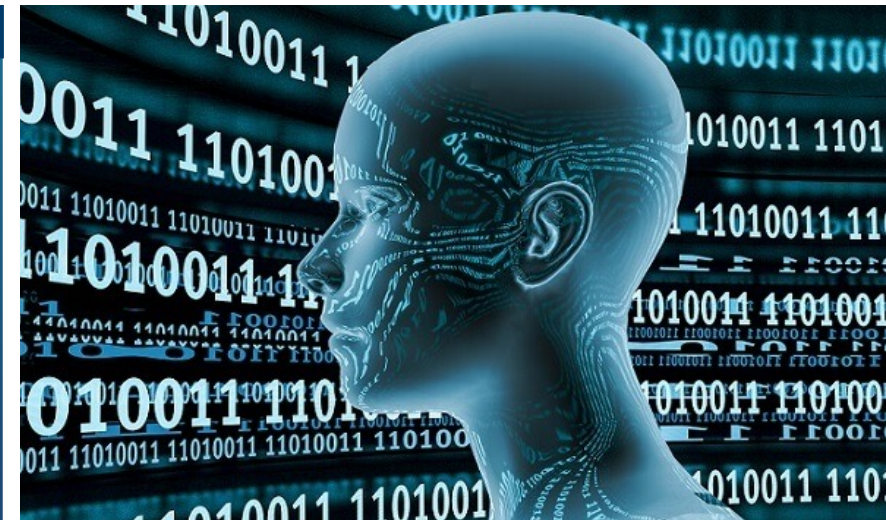
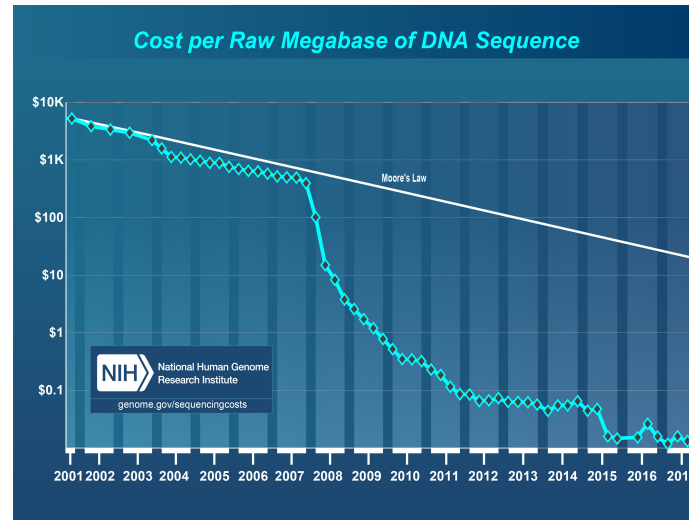
- How can we find evidence of **resistance** in genomic data?
- How can we find evidence of **susceptibility** in genomic data?
 - Is this just the absence of (known) resistance determinants? Is that enough?
 - Works well in certain cases: MRSA, VRE, TB
 - But... these are corner cases
 - Consider: 13-68% of CRE^{2,3}, and >95% of carba-R *Pseudomonas*³, do NOT have carbapenemases

²Cerqueira GC *et al*, *PNAS* 2017;114:1135

³Woodworth KR *et al*, *MMWR* 2018;67:396

Genomics and AST

- How can we find evidence of **resistance** in genomic data?
- How can we find evidence of **susceptibility** in genomic data?
 - Is this just the absence of (known) resistance determinants? Is that enough?
 - Can't we just sequence everything and use machine learning / AI?
 - *Correlates vs surrogates*
 - *Agnostic vs mechanistic*



Genotypic vs phenotypic approaches for antibiotic susceptibility testing (AST)



• Phenotypic AST



- Long the gold standard
- Detects **susceptibility** - "answers the key question"
 - Also risks of errors - eg carbapenemases: inoculum effect, heteroresistance
- Speed?
- Antibiogram as (very) low-res method to infer transmission

• Genotypic AST

- Capitalizes on WGS revolution
- Predicts **resistance** - "by proxy" or "by correlate"
 - Risk of errors from undertraining, unexpected diversity, new mechanisms
- Speed? Cost? Implementation?
- "Collateral info": potential for built-in molecular epidemiology

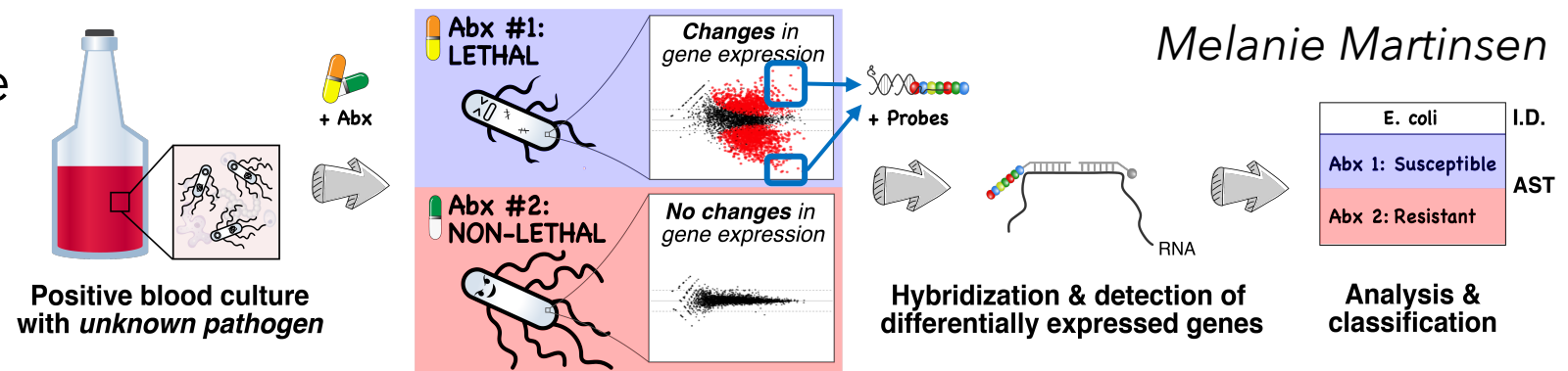
What if we could do both in a single assay?

A new approach to AST through RNA detection: *transcription as phenotype*

- RNA **sequence** → **genotype**
- RNA **abundance** → **phenotype**
 - Transcriptional changes are among the earliest adaptations to stress

- **Postulate:** **susceptible** and **resistant** strains will exhibit **differential gene expression upon antibiotic exposure**

- Rapid (minutes)
- Agnostic to resistance mechanism

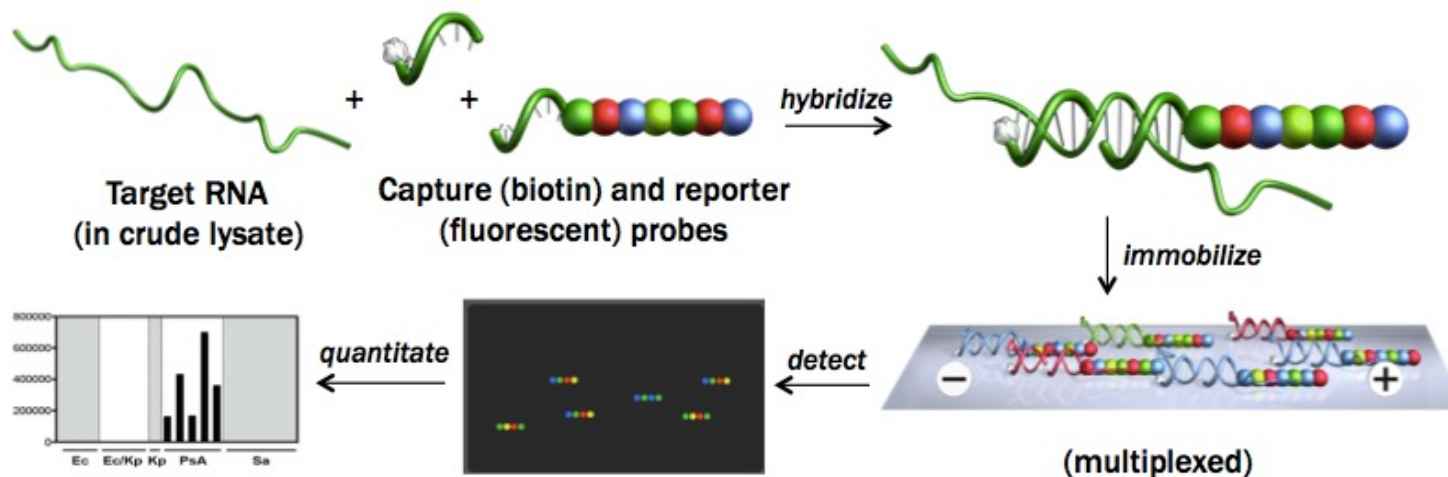


GoPhAST-R: Genotypic + Phenotypic AST through RNA detection

RNA signatures: an early, readily measurable distress signal

- RNA **sequence** → **genotype**
- RNA **abundance** → **phenotype**
 - Transcriptional changes are among the earliest adaptations to stress

- Enabling technology: **NanoString**



- **Multiplexable** (hundreds of transcripts in "one pot")
- **Quantitative** over 3-4 orders of magnitude
- Total assay time ~hours (hands-on time: ~minutes)
- No enzymology = direct from crude lysates
- Benchtop instrument for detection

Workflow: defining transcriptional signatures

Drug exposure



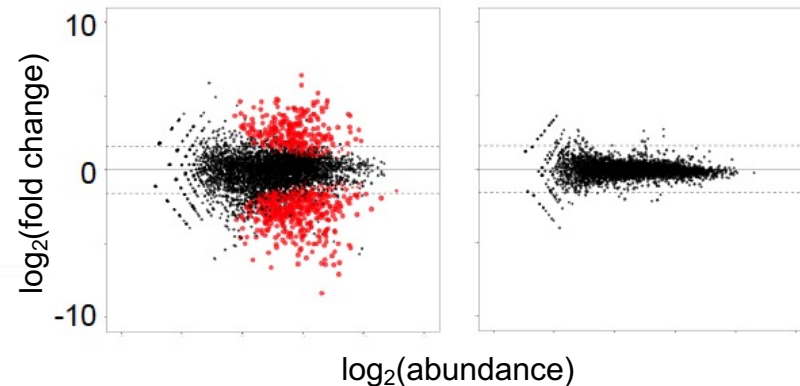
clinical isolates
(susc, res)
+/- antibiotic
(at clinical breakpoint)

Expression analysis
(RNA-Seq)

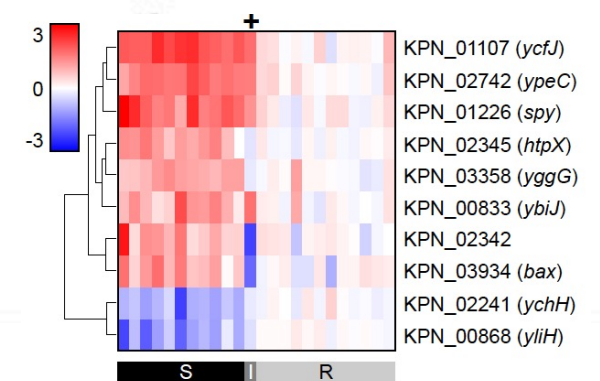


susc

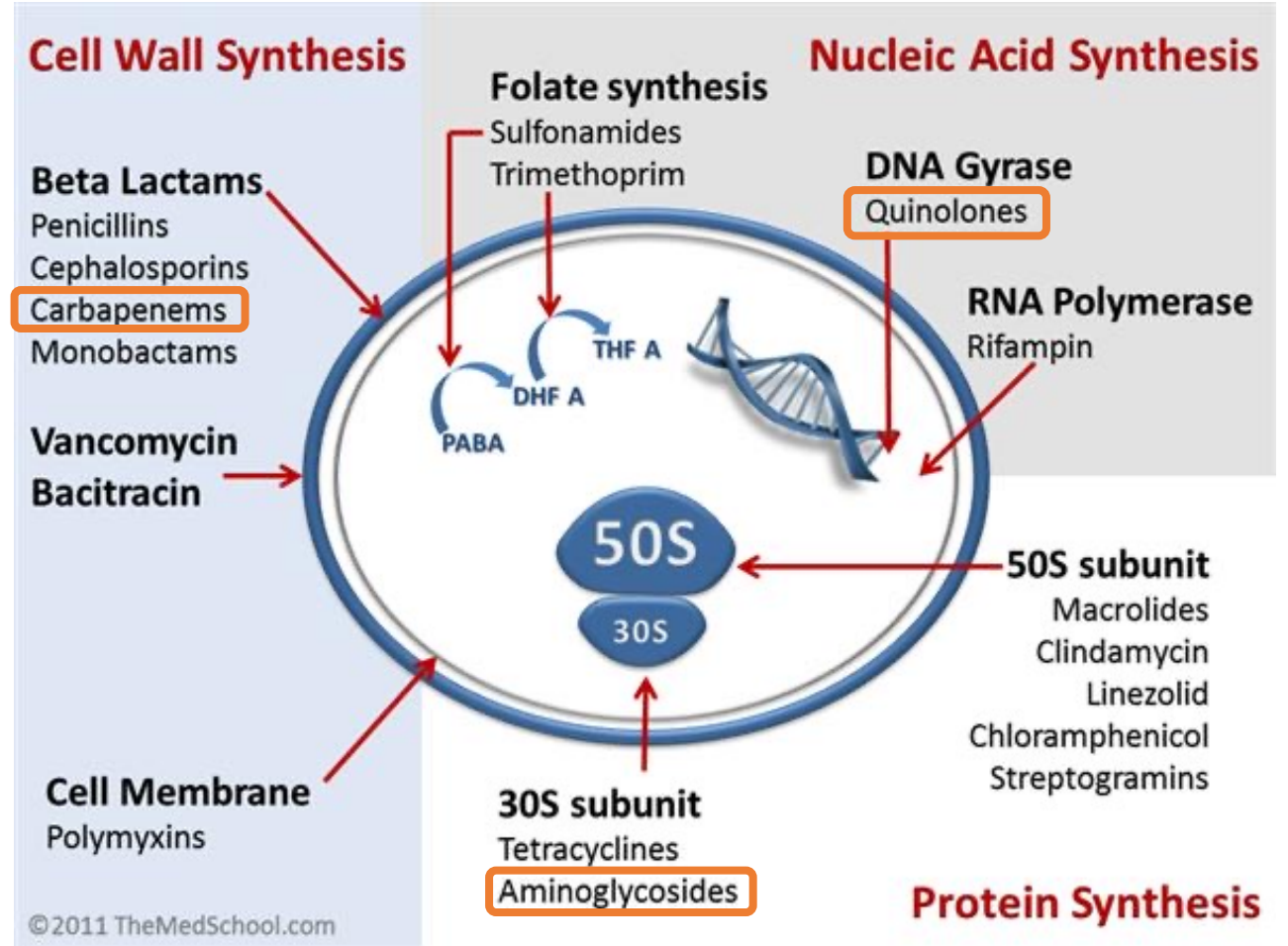
res



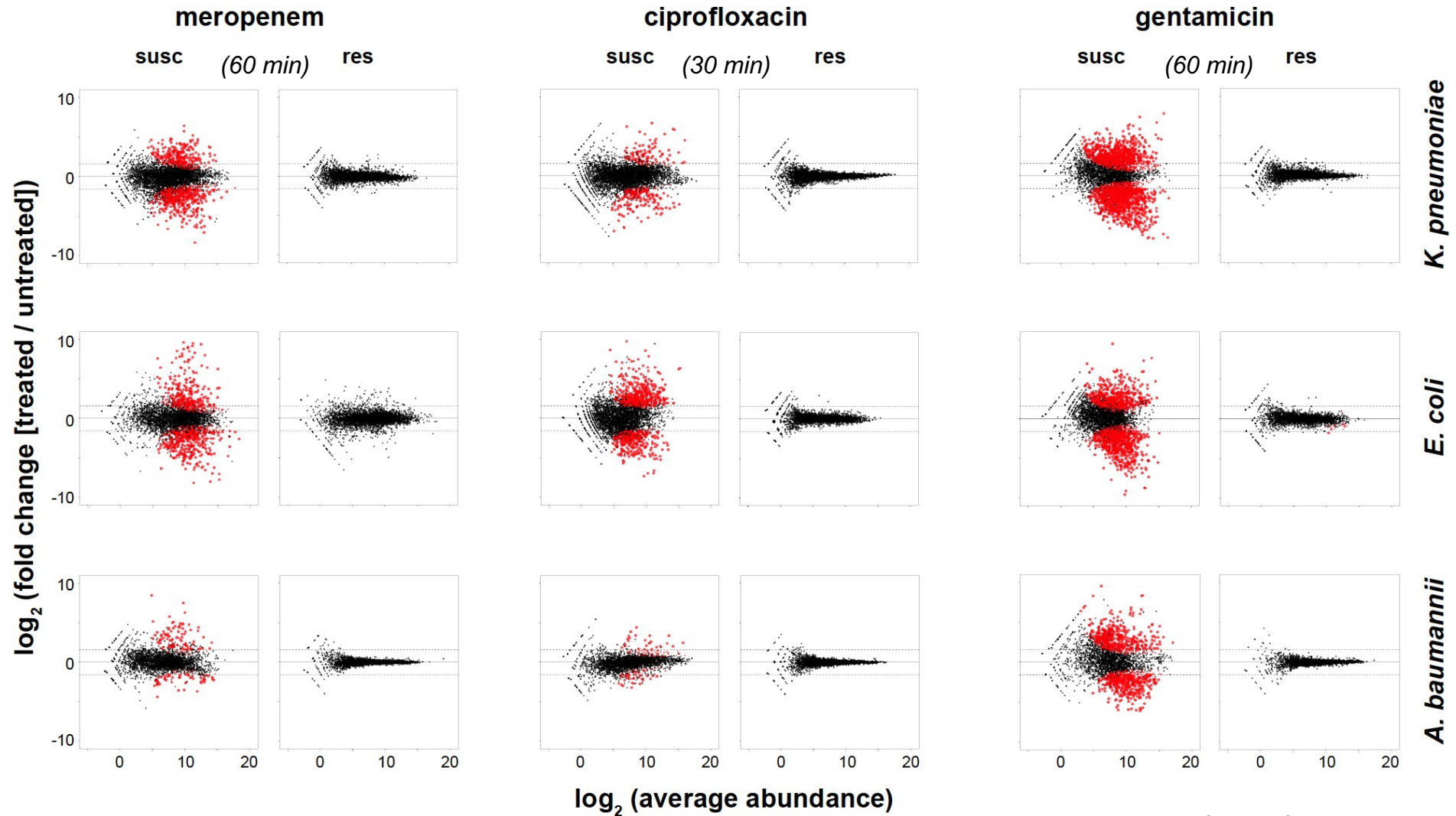
Signature validation
(multiplexed RNA detection)



Application: MDRO GNRs, multiple antibiotic classes



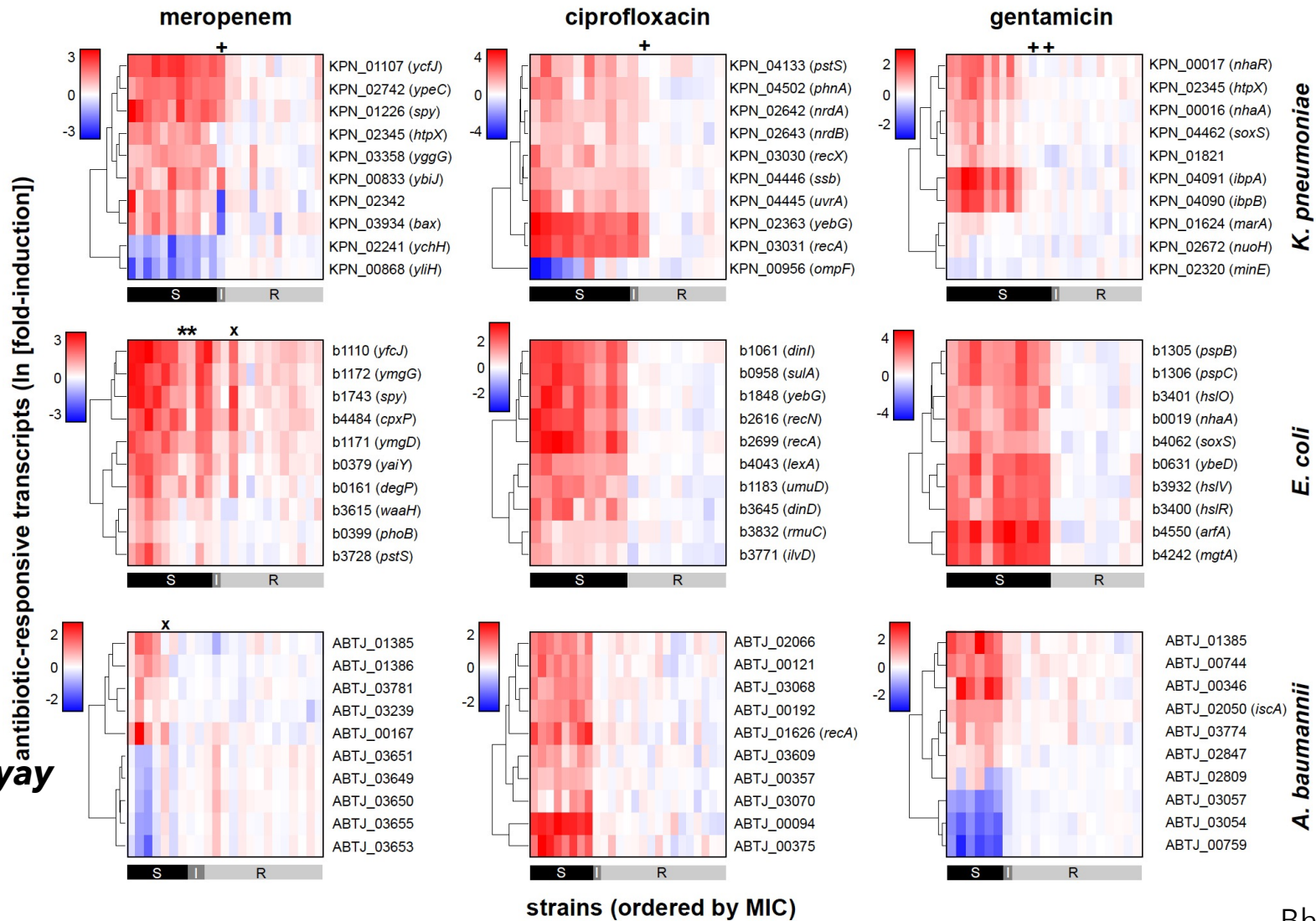
RNA-Seq: S and R strains respond differently to antibiotic exposure



● $p_{adj} < 0.05$

J. Livny
J. Liu
D. Hung

GoPhAST-R: a small subset of transcripts predict AST



(top 10 antibiotic-responsive transcripts, measured by NanoString hybridization assay)

N. Bandyopadhyay

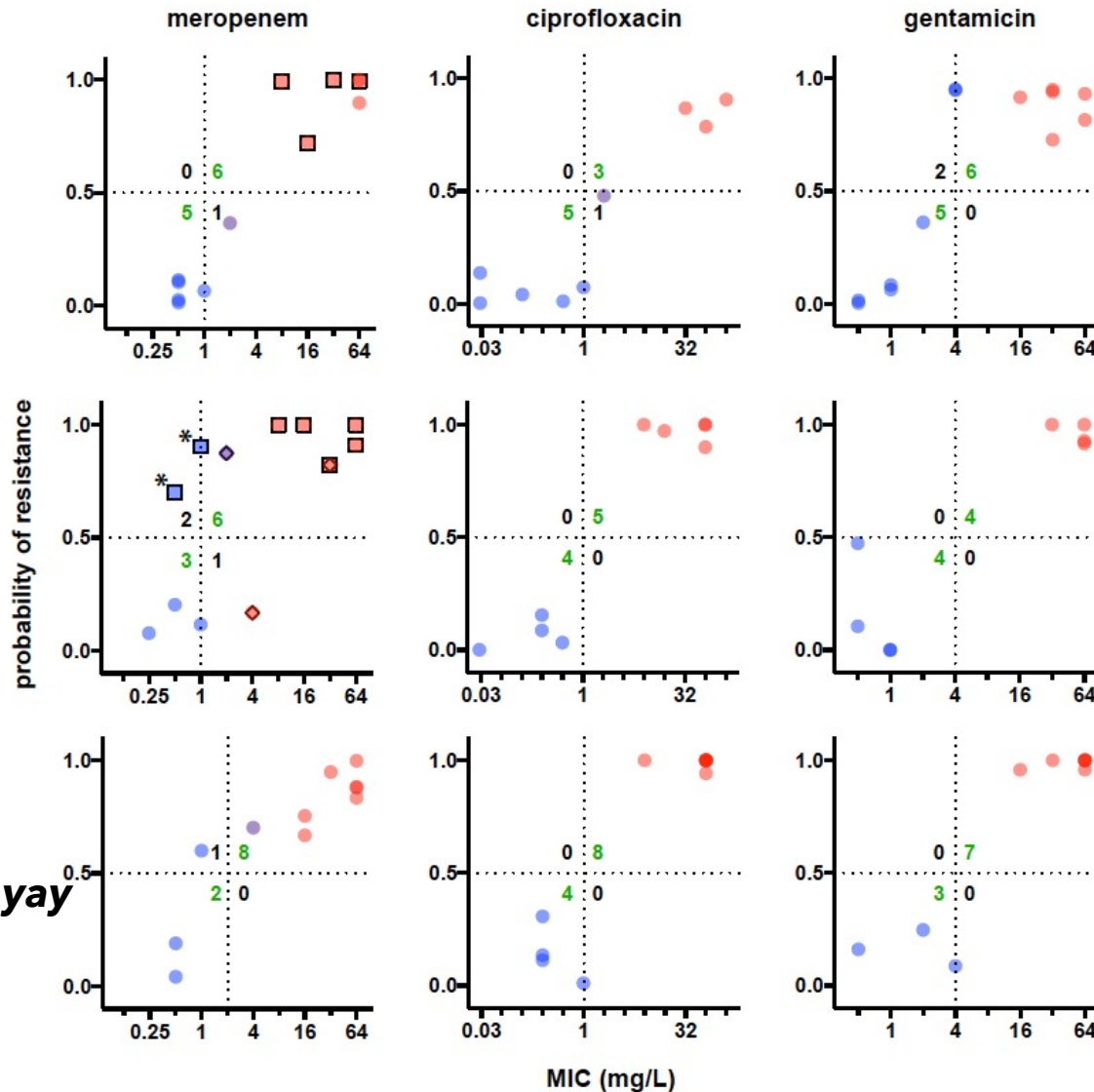
J. Livny

N. Shores

S. Son

D. Hung

GoPhAST-R: machine learning predicts susceptibility from NanoString data



Input: NanoString data, top 10 antibiotic-responsive transcripts

Model: random forest

Output: probability of resistance

		Reference (MIC)		
		Susc	Intd	Res
Prediction (GoPhAST-R)	Susc	97	5	1
	Non-susc (I or R)	6	23	107

Categorical agreement: 227/239 (95%)

- 3 discrepancies clearly due to errors w/ gold standard
- 8 of remaining 9 "missed close" (variability in gold standard too)

N. Bandyopadhyay

J. Livny

N. Shoresh

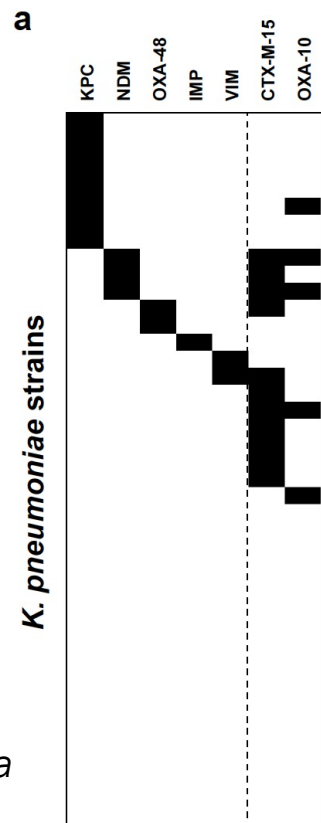
S. Son

D. Hung

GoPhAST-R: simultaneous resistance gene detection enhances assay

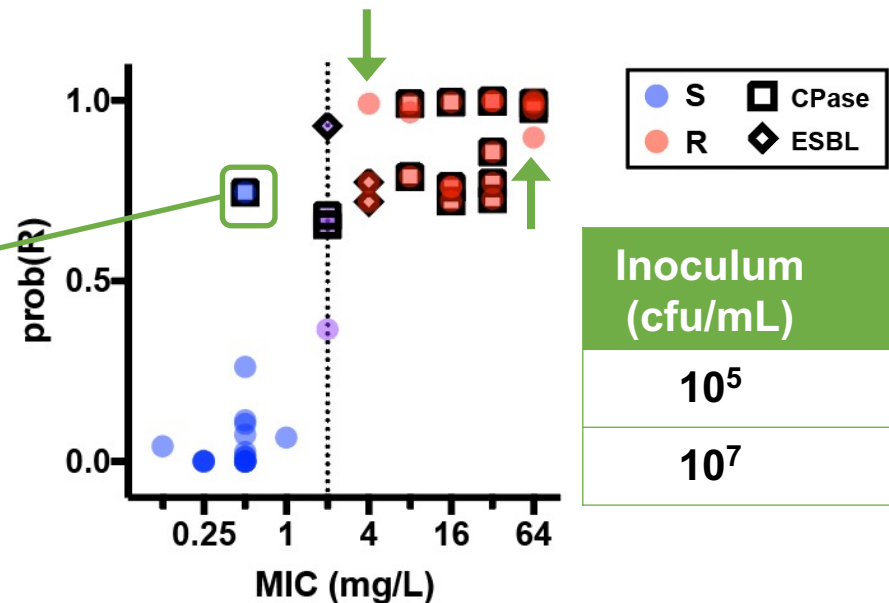
- Simultaneous detection of 5 common carbapenemases:
- Overlay on phenotypic AST

resistance gene content



WGS: present
 absent

Detected 67/67 carbapenemases



Inoculum (cfu/mL)	MIC (mg/L)
10 ⁵	0.5
10 ⁷	32

- Uncover "hidden" carbapenem-R
- Flag unexplained resistance
- Suggest alternative abx (avibactam; aztreonam)
- Facilitate molecular epidemiology

J. Livny

P. Ma

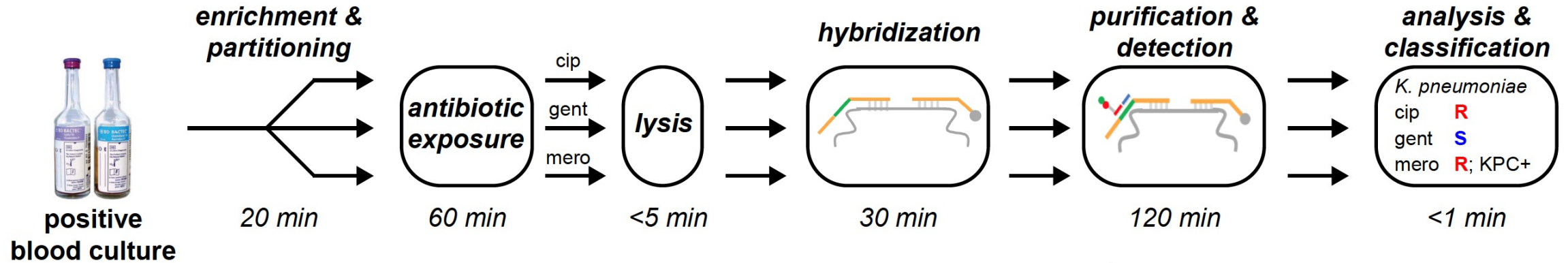
S. Son

G. Cerqueira

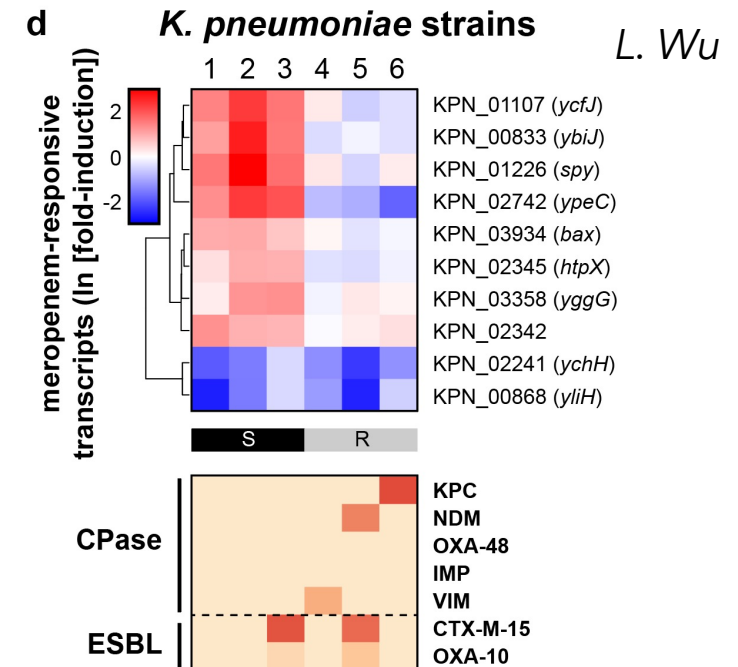
A. Earl

D. Hung

GoPhAST-R: proposed workflow

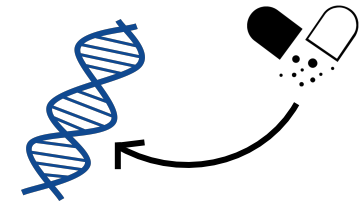
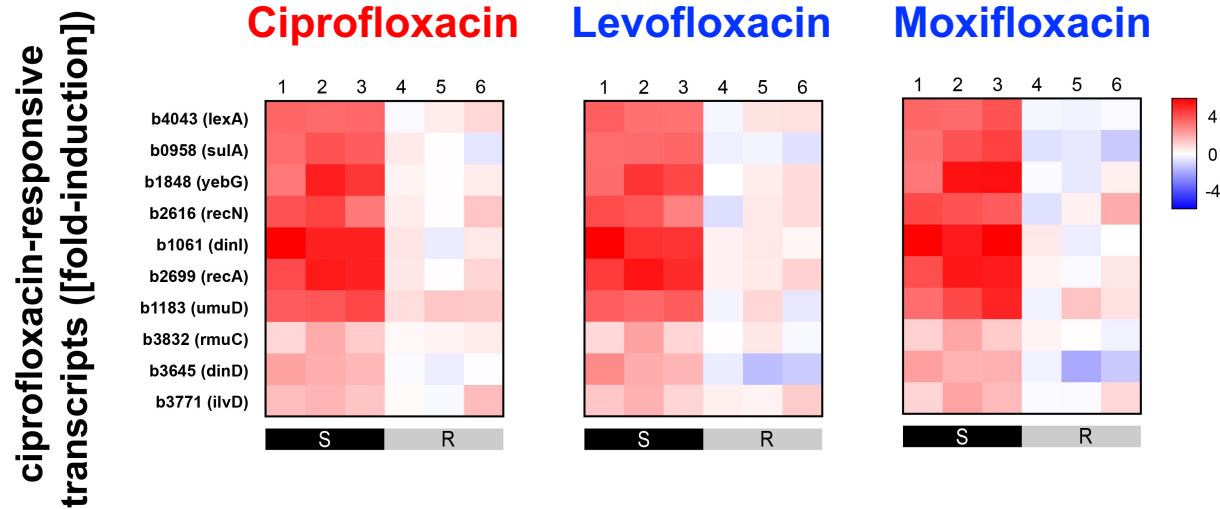


- **AST** in <4 hours (<30 min hands-on)
 - vs broth microdilution = 28 hrs from positive BCx
- Success direct from blood cultures
 - 71/72 (99%) correct from spiked BCx
 - 8/8 correct for real BCx
- Genotype: built-in carbapenemase assay
- RNA-Seq data on ~50 bug-drug pairs



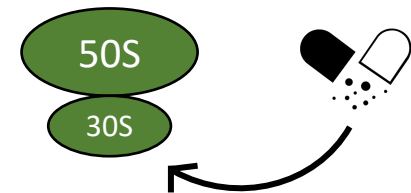
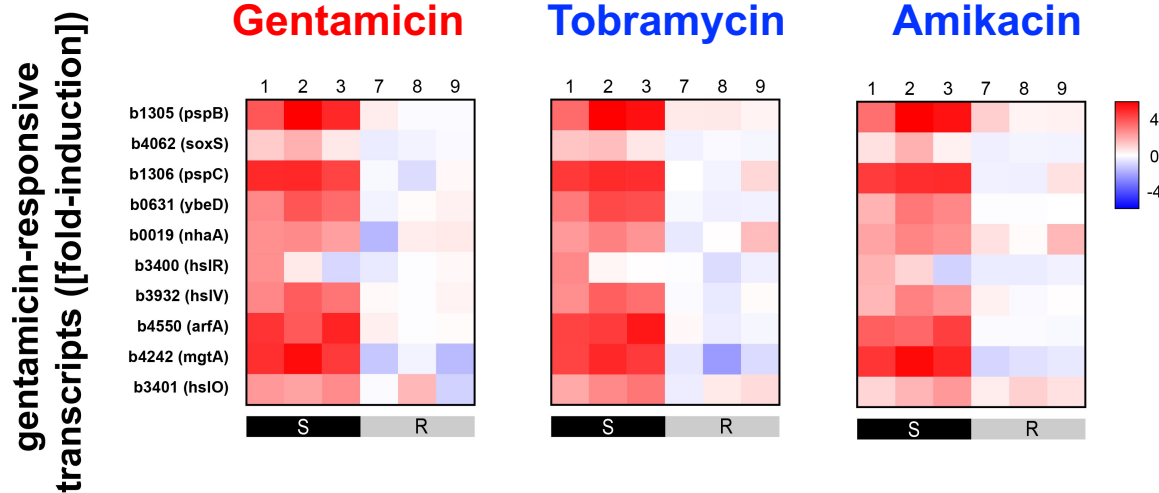
RNA signatures generalize across fluoroquinolones & aminoglycosides

RNA-Seq → NanoString

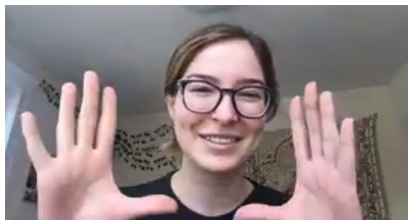


DNA gyrase inhibitors

NanoString only



protein synthesis inhibitors



Melanie Martinsen

Strains (ordered by CLSI classification)

Martinsen et al, AAC 2021

RNA signatures generalize across beta-lactams

Most narrow

Most broad

meropenem-responsive transcripts ([fold-induction])

Ampicillin

Cefazolin

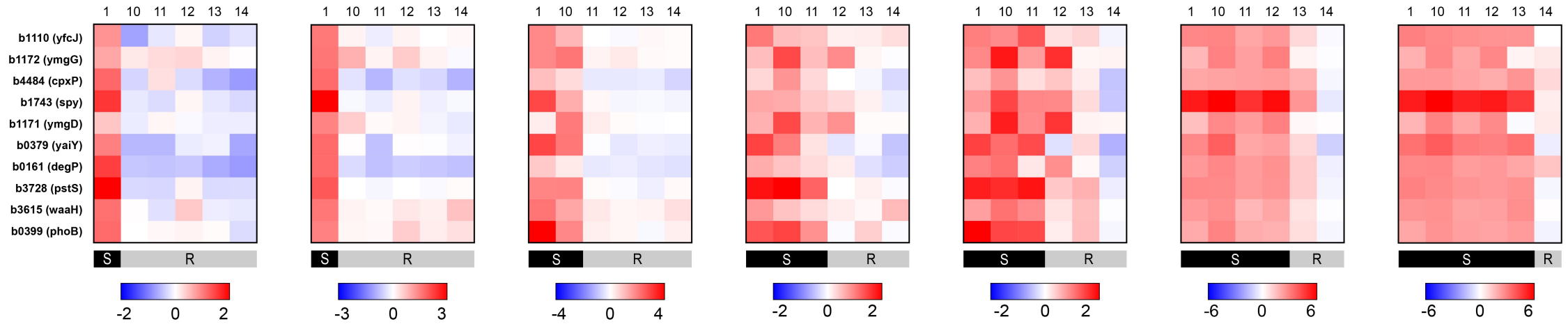
Ceftriaxone

Aztreonam

Piperacillin/
tazobactam

Ertapenem

Meropenem

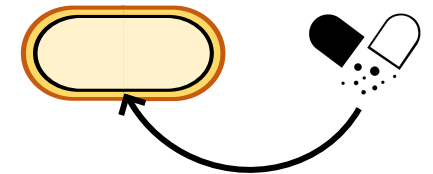


Strains (ordered by CLSI classification)



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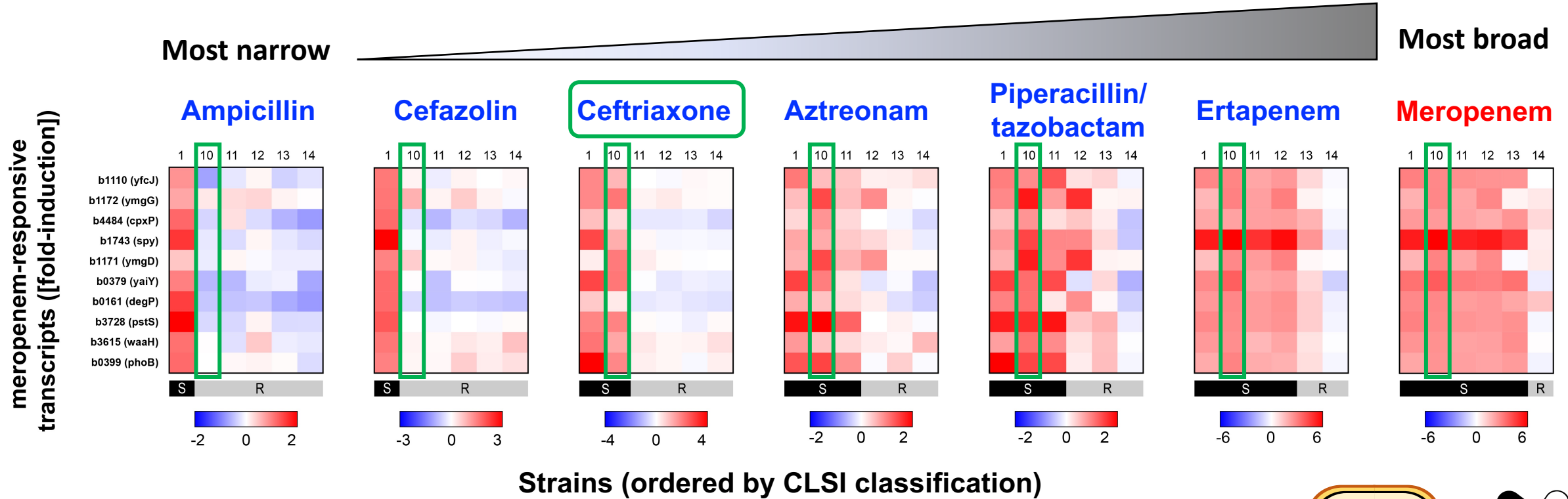
Alex Jaramillo Cartagena



cell wall synthesis inhibitors

Martinsen et al, AAC 2021

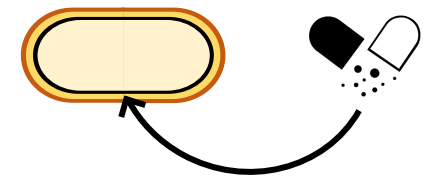
RNA signatures generalize across beta-lactams



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cell wall synthesis inhibitors

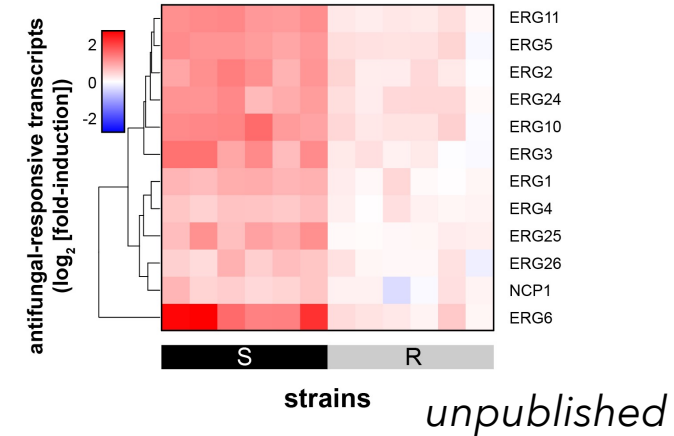
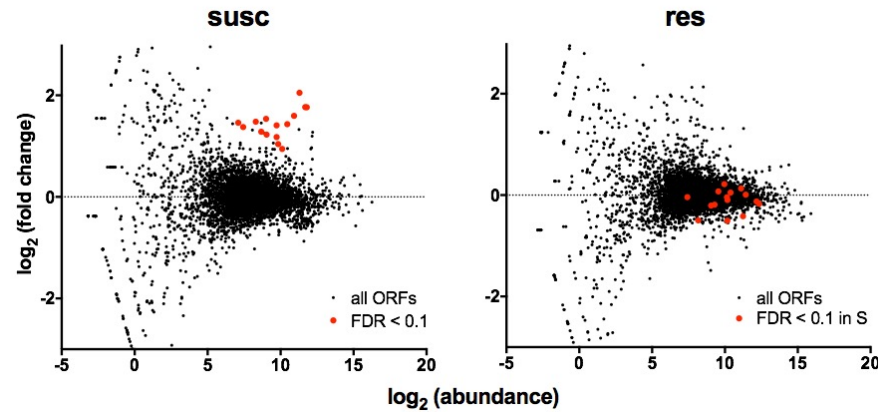
Martinsen et al, AAC 2021

Beyond typical bacteria: slow-growing species still transcribe fast

- Pilot fungal transcriptional signatures enable rapid AST on the same NanoString platform:

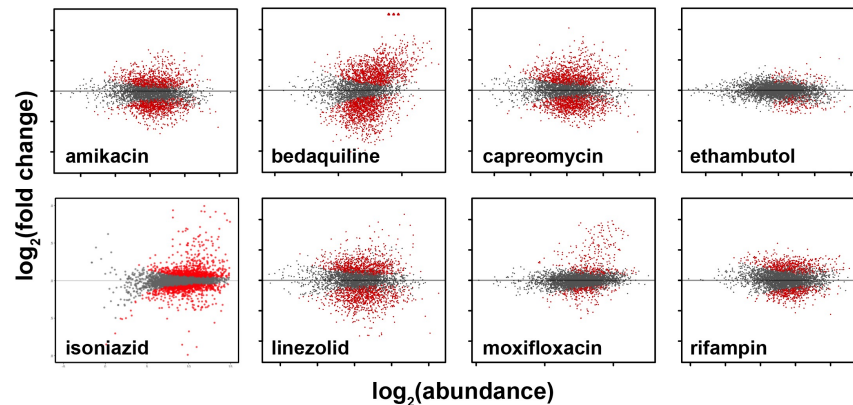
Melanie Martinsen
 Eleanor Young
 Jose Muñoz
 Poppy Sephton-Clark
 Michelle Matzko
 Christina Cuomo

Candida albicans
 + fluconazole:



- *Mycobacterium* show transcriptional signatures in hours:

M. tuberculosis +
 various abx:



in prep

with A. van den
 Bossche, P. Ceysens
 (Sciensano, Belgium)

Summary: RNA-based detection for rapid bacterial ID and AST

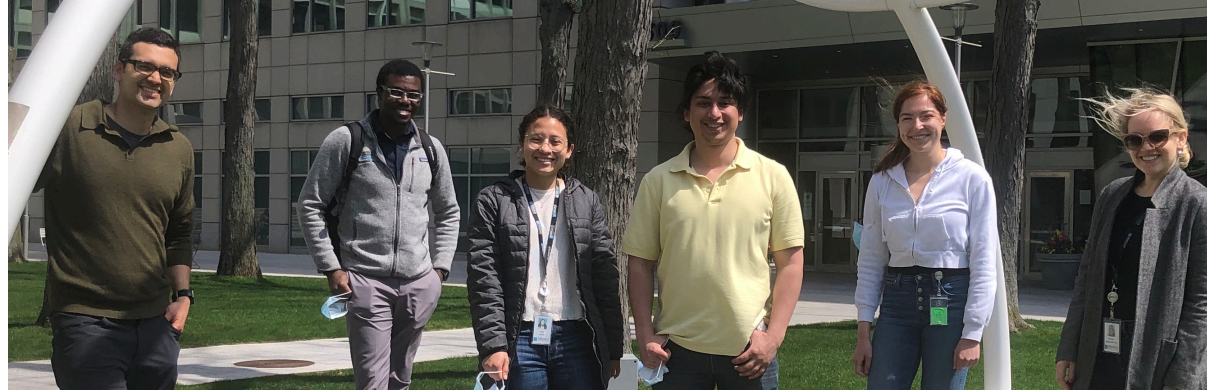
- **GoPhAST-R:** antibiotic-induced transcriptional signatures are a fast, accurate, **phenotypic** measure of **antibiotic susceptibility**
 - Sensitive to $<10^5$ bacteria; assay time ~4 hrs
 - Clinical pilot on BCx underway
- Simultaneous detection of key genetic resistance determinants enhances AST accuracy, value
- Accuracy likely to improve with further training/testing
- Success in fungi, mycobacteria = possible **pan-microbial** approach to AST (work in progress)
- Goals: faster, cheaper, more sensitive, & more deployable assay
- What biology underlies these responses?
 - Shared pathways within, among classes? Adaptation? Signs of struggle?



Acknowledgments



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 - Kyra Taylor
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 - Lidan Wu
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 - Sophie Son
 - Noam Shores



Broad Fungal Genomics

- **Christina Cuomo**
- **Poppy Sephton-Clark**
- Jose Muñoz

Broad Bacterial Genomics

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- Abigail Manson

MGH

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BWH

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- Tulip Jhaveri
- Sanjat Kanjilal

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- Kim Musser

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• MGH ID Division